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APPLICATION NO.	FILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO.
10/630,357	07/30/2003	Sammy S. Datwani	100/17201	9263
	7590 08/16/2007 E SCIENCES, INC.		EXAMINER	
605 FAIRCHIL	LD DRIVE		WALLENHORST, MAUREEN	
MOUNTAIN	/IEW, CA 94043-2234		ART UNIT PAPER NUMBER	
			1743	
			MAIL DATE	DELIVERY MODE .
			08/16/2007	PAPER

Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

		Application No.	Applicant(s)			
		10/630,357	DATWANI ET AL.			
	Office Action Summary	Examiner	Art Unit			
		Maureen M. Wallenhorst	1743			
Period fo	The MAILING DATE of this communication apport Reply	pears on the cover sheet with	the correspondence address			
A SH WHIC - Exte after - If NC - Failu Any	ORTENED STATUTORY PERIOD FOR REPLY CHEVER IS LONGER, FROM THE MAILING Donsions of time may be available under the provisions of 37 CFR 1.1 SIX (6) MONTHS from the mailing date of this communication. Operiod for reply is specified above, the maximum statutory period were to reply within the set or extended period for reply will, by statute reply received by the Office later than three months after the mailing ed patent term adjustment. See 37 CFR 1.704(b).	ATE OF THIS COMMUNICA 36(a). In no event, however, may a reply will apply and will expire SIX (6) MONTHS c. cause the application to become ABAN	TION. be timely filed from the mailing date of this communication.			
Status						
1) 又	Responsive to communication(s) filed on 24 Ju	ulv 2007.	·			
	This action is FINAL . 2b)⊠ This action is non-final.					
3)						
	closed in accordance with the practice under E					
Disposit	ion of Claims		•			
4)⊠	Claim(s) <u>75,77-85,87-90 and 92-102</u> is/are per	nding in the application.				
	4a) Of the above claim(s) is/are withdraw					
	Claim(s) is/are allowed.					
6)⊠	Claim(s) <u>75, 77-85, 87-90, 92-102</u> is/are rejected.					
7)	•					
8)[Claim(s) are subject to restriction and/o	r election requirement.	·			
Applicat	ion Papers					
9)[The specification is objected to by the Examine	er.				
	The drawing(s) filed on is/are: a) acc		the Examiner			
•	Applicant may not request that any objection to the	· · · · · · · · · · · · · · · · · · ·				
	Replacement drawing sheet(s) including the correct	tion is required if the drawing(s)	is objected to. See 37 CFR 1.121(d).			
11)	The oath or declaration is objected to by the Ex	caminer. Note the attached O	ffice Action or form PTO-152.			
Priority (ınder 35 U.S.C. § 119					
	Acknowledgment is made of a claim for foreign ☐ All b)☐ Some * c)☐ None of:	priority under 35 U.S.C. § 1	19(a)-(d) or (f).			
	1. Certified copies of the priority document	s have been received.				
	2. Certified copies of the priority document	s have been received in App	lication No			
	3. Copies of the certified copies of the prior	rity documents have been re	ceived in this National Stage			
	application from the International Bureau	• • • • • • • • • • • • • • • • • • • •				
* 5	See the attached detailed Office action for a list	of the certified copies not rec	ceived.			
			•			
Attachmen	t(s)					
1) Notice	ce of References Cited (PTO-892)		mary (PTO-413)			
	ce of Draftsperson's Patent Drawing Review (PTO-948) mation Disclosure Statement(s) (PTO/SB/08)		lail Date mal Patent Application			
	er No(s)/Mail Date	6) Other:				

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1. A request for continued examination under 37 CFR 1.114, including the fee set forth in 37 CFR 1.17(e), was filed in this application after final rejection. Since this application is eligible for continued examination under 37 CFR 1.114, and the fee set forth in 37 CFR 1.17(e) has been timely paid, the finality of the previous Office action has been withdrawn pursuant to 37 CFR 1.114. Applicant's submission filed on July 24, 2007 has been entered.

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- 2. This application currently names joint inventors. In considering patentability of the claims under 35 U.S.C. 103(a), the examiner presumes that the subject matter of the various claims was commonly owned at the time any inventions covered therein were made absent any evidence to the contrary. Applicant is advised of the obligation under 37 CFR 1.56 to point out the inventor and invention dates of each claim that was not commonly owned at the time a later invention was made in order for the examiner to consider the applicability of 35 U.S.C. 103(c) and potential 35 U.S.C. 102(e), (f) or (g) prior art under 35 U.S.C. 103(a).
- 3. The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:
 - (a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negatived by the manner in which the invention was made.
- 4. The factual inquiries set forth in *Graham* v. *John Deere Co.*, 383 U.S. 1, 148 USPQ 459 (1966), that are applied for establishing a background for determining obviousness under 35 U.S.C. 103(a) are summarized as follows:
 - 1. Determining the scope and contents of the prior art.
 - 2. Ascertaining the differences between the prior art and the claims at issue.
 - 3. Resolving the level of ordinary skill in the pertinent art.

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- 4. Considering objective evidence present in the application indicating obviousness or nonobviousness.
- 5. Claims 75, 77-78, 82-85, 87-90 and 92-98 are rejected under 35 U.S.C. 103(a) as being unpatentable over Wolk et al (US Patent no. 6,620,625, submitted in the Information Disclosure Statement filed on March 1, 2004).

Wolk et al teach of an ultra high throughput sampling and analysis system used for sampling large numbers of different materials from surfaces of substantially planar library storage components. The system comprises a substrate 704 that has a large number of discrete quantities of different test compounds of reagents removably immobilized thereon. Removably immobilized means that the sample materials are present upon the sample substrate in an immobilized format (confined in a discrete region), but are removable from the substrate through appropriate action. The samples are deposited on the substrate in an array and dried. The samples are removable by dissolving them in a fluid and pulling the fluid off the sample substrate. The sample fluids may be coupled to the substrate through ionic, hydrophobic or hydrophilic interactions, and covalent interactions, which are severable by exposing the substrate to an appropriate environment such as a certain buffer solution, thermal environment, etc. Hydrophobic barriers surrounding hydrophilic regions may be present on the substrate to separate the individual reagent spots, or vice-versa. The substrate can be made out of a nonporous material such as glass, silicon or a polymer. The substrate surface may have a coating thereon such as a glass substrate coated with a silane material or a surface containing a polymeric or metallic coating. The substrates can have a pattern thereon such as a honeycomb pattern with uniformly spaced pores for holding the reagent spots. The reagent spots can comprise proteins, peptides, nucleic acids and the like. The compounds that are spotted onto the substrate comprise

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in addition to the particular compound at least one excipient material that enhances one or more of the deposition and/or solubilization of the compound in an appropriate solubilization liquid. Examples of excipients include polymers such as PEG, detergents, sugars and solvents such as DMSO. See lines 42-65 in column 13 and lines 12-17 in column 27 of Wolk et al. An automated system is used to position the substrate in relation to a microfluidic device having an external sampling capillary element 106 attached thereto. The substrate and microfluidic device can be positioned on a platform that is movable in the X-Y-Z direction so as to move the array and microfluidic device relative to one another. Wolk et al teach that the substrate may contain alignment marks thereon that are marks on the array that correspond to a certain position on the array. The alignment marks may contain material that makes them fluoresce and be opaque. An imaging system is used to locate the alignment marks on the substrate in order to position the capillary element 106 of the microfluidic device above one of the reagent spots on the array. Once a reagent spot is located by means of the alignment marks, the capillary element serves to dispense a solvent or buffer solution to the reagent spot in order to dissolve the reagent spot. The dissolved reagent is then aspirated by the capillary element 106 so as to enter into the microfluidic device for analysis. See figures 1, 6 and 7a, lines 9-62 in column 2, lines 36-67 of column 4, lines 8-30 in column 5, lines 32-65 in column 9, lines 34-64 in column 11, lines 24-43 in column 12, lines 42-65 in column 13, lines 19-44 in column 19, lines 15-67 in column 20, lines 1-11 in column 21 and lines 12-28 in column 27 of Wolk et al.

Wolk et al fail to teach that the alignment marks on the substrate comprise both a dye and a substantially water insoluble polymer excipient. However, it would have been obvious to one of ordinary skill in the art at the time of the instant invention to realize or recognize that the

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alignment marks on the substrate taught by Wolk et al contain a dye therein since Wolk et al teach that 1) the alignment marks must be visually detectable by an imaging system, thus making it highly likely that a dye is contained in the marks, and 2) the marks may exhibit fluorescent properties and be opaque (see lines 60-67 in column 20 of Wolk et al), thus suggesting that the marks contain a dye therein. It also would have been obvious to one of ordinary skill in the art at the time of the instant invention to include in the alignment marks taught by Wolk et al a substantially water insoluble polymer excipient since Wolk et al disclose that such polymer excipients are advantageously combined with the reagents that are spotted onto the substrate in an array so as to enhance the binding and deposition of the reagents to the substrate, and therefore, one of skill in the art would have found it obvious to also combine the material of the alignment marks with a similar type of polymer excipient material in order to help enhance the binding and deposition of the alignment mark material onto the substrate. It also would have been obvious to one of ordinary skill in the art to use a polymer excipient that is water insoluble in combination with the alignment mark materials taught by Wolk et al so as to prevent the alignment marks from being washed away from the surface of the substrate upon contact of the substrate with aqueous liquids that are used to solubilize the reagents of the array. Regarding claims 77-78, 87-90 and 92-95, it would have been obvious to one of ordinary skill in the art to use the dye and excipient materials and concentrations as recited in these claims for the alignment marks taught by Wolk et al since the determination of an optimal material and concentration of the material for a particular use is easily determined by routine experimentation.

6. Claims 79-81 and 99-102 are rejected under 35 U.S.C. 103(a) as being unpatentable over Wolk et al in view of Wagner et al (US Patent no. 6,475,809). For a teaching of Wolk et al, see

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previous paragraphs in this Office action. Wolk et al fail to teach that the substrate on which the array of reagents is spotted comprises a self-assembled monolayer formed at an interface on the substrate surface.

Wagner et al teach of protein arrays for high-throughput screening. The device comprises a substrate having a plurality of spots of proteins arranged in discrete, known locations. The proteins are spaced from one another in rows and columns from about 50 nm to about 500 micrometers apart. The substrate can be made of glass, silicon, silica, a polymer or a metal. In a preferred embodiment, the proteins are attached onto a self-assembled monolayer that is attached to an interface on the substrate. The monolayer contains molecules of the formula X-R-Y, wherein R is a spacer, X is a functional group that binds R to the surface, and Y is a functional group for binding proteins onto the monolayer. The array may also comprise a coating between the substrate and the monolayer. The coating is applied to the substrate using techniques such as physical vapor deposition or chemical vapor deposition. The coating may comprise a metal film such as aluminum, chromium, gold or silver. The coatings may require an adhesion layer between the coating and the substrate such as a layer of titanium. The deposition of the coating on the substrate is done prior to the formation of patches of proteins thereon. Monolayer-compatible surface coatings may be fabricated in a pattern onto the substrate using photolithography, chemical etching or micro molding. Protein patches are then formed in the openings of the pattern. Diffusion boundaries between the openings of the pattern are formed as walls of substrate material or photo resist. The walls of the pattern are used to separate the protein patches from one another. In a preferred embodiment, the patches or openings are separated from one another by surfaces of the substrate free of monolayer of the form X-R-Y.

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Alternatively, non-bioreactive monolayers with a different wettability may be used to separate patches of proteins from one another. See Figure 2 in Wagner et al that depicts a coating 5 on a substrate 3, an adhesion interlayer 6 and a monolayer 7. The R group in the self-assembled monolayer comprises a hydrocarbon chain of from about 1 to 200 carbons long. Preferably the R is an alkyl chain having about 8 to 22 carbons, such as a straight alkane. X is any group that affords chemisorption or physisorption of the monolayer onto the surface of the substrate (or the coating if present). When the substrate or coating is a metal, X is chosen to be a disulfide, a sulfide, a thiol or a trivalent phosphorus compound. This embodiment is preferred when the coating or substrate is gold or silver. If the substrate is a glass or silicon material, X is preferably a silane material such as a monohalosilane, a dihalosilane, etc. When the surface of the substrate is composed of a metal oxide, X is preferably a carboxylic acid. The component Y of the monolayer is a functional group responsible for binding a protein onto the monolayer. The Y group may form a covalent or noncovalent linkage with the protein. Possible Y groups include -OH, NH2, -COOH, -PO4, SO3, etc. Wagner et al teach that a method of using the protein array attached to a self-assembled monolayer on a substrate involves screening the proteins for their ability to interact with a component of a fluid sample. The method of use comprises delivering a fluid sample to the array, and detecting the interaction of the component with the immobilized protein of each patch or spot. See figures 1, 3, 4-7, lines 57-67 in column 7, lines 1-67 in column 8, lines 1-67 in column 9, lines 1-63 in column 10, lines 25-35 in column 11, lines 3-39 in -column 12, and lines 40-46 in column 17 of Wagner et al.

Based upon the combination of Wolk et al and Wagner et al, it would have been obvious to one of ordinary skill in the art at the time of the instant invention to attach the array of

reagents taught by Wolk et al onto a self-assembled monolayer formed at an interface of the substrate since Wolk et al teach that the reagents must be reversibly attached to the substrate in an ordered array, and Wagner et al teach that self-assembled monolayers attached to an interface of a substrate allows reagent materials such as proteins to be reversibly attached to the substrate in an organized and ordered array without the broad and irregular spreading of the protein spots into one another. Thus, the provision of a self-assembled monolayer at an interface on the substrate taught by Wolk et al would allow for a more consistent and uniform surface to receive the reagents of the array without any cross contamination.

7. Applicant's arguments filed July 24, 2007 have been fully considered but they are not persuasive.

The previous objections to the claims made in the Office action mailed on April 24, 2007 have been withdrawn in view of Applicants' amendments to the claims. The previous rejections of the claims under 35 USC 103 as being obvious over the combination of references to Wolk et al, Ezrielev et al and Wagner et al have been withdrawn in view of Applicants' amendments to the claims and persuasive arguments. However, upon further consideration, the claims are now rejected under 35 USC 103 as being obvious over either Wolk et al or obvious over Wolk et al in view of Wagner et al. For a discussion of why the claims are now newly rejected using these references and the Examiner's reasoning, see previous paragraphs in this Office action.

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8. Any inquiry concerning this communication or earlier communications from the

examiner should be directed to Maureen M. Wallenhorst whose telephone number is 571-272-

1266. The examiner can normally be reached on Monday-Thursday from 6:00 AM to 4:30 PM.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's

supervisor, Jill Warden, can be reached on 571-272-1267. The fax phone number for the

organization where this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent

Application Information Retrieval (PAIR) system. Status information for published applications

may be obtained from either Private PAIR or Public PAIR. Status information for unpublished

applications is available through Private PAIR only. For more information about the PAIR

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system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free).

Maureen M. Wallenhorst Primary Examiner

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mmw

August 9, 2007

Naureen M. Wallenhorst
MAUREEN M. WALLENHORST
PRIMARY EXAMINER
GROUP 1999 (700